

Package ‘dPCP’

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Title Automated Analysis of Multiplex Digital PCR Data

Version 1.0.4

Description The automated clustering and quantification of the digital PCR data is based on the combination of 'DBSCAN' (Hahsler et al. (2019) <[doi:10.18637/jss.v091.i01](https://doi.org/10.18637/jss.v091.i01)>) and 'c-means' (Bezdek et al. (1981) <[doi:10.1007/978-1-4757-0450-1](https://doi.org/10.1007/978-1-4757-0450-1)>) algorithms.

The analysis is independent of multiplexing geometry, dPCR system, and input amount.

The details about input data and parameters are available in the vignette.

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centers_data	<i>Prediction of clusters centroid position</i>
--------------	---

Description

This function calculates the coordinates of all clusters centroid.

Usage

```
centers_data(sample.subquality, sample.table, referenceDB)
```

```
## S3 method for class 'centers_data'
plot(x, ..., sample = "all")
```

Arguments

sample.subquality	an object of class read_sample, inherited from read_sample .
sample.table	object of class sample_table, inherited from read_sampleTable .
referenceDB	an object of class reference_dbscan, inherited from reference_dbscan
x	an object of class centers_data
...	Arguments to be passed to methods
sample	'all' to show all samples, or a numeric vector indicating the row number of samples in the sample table.

Value

An object of class `centers_data` containing a sublist for each sample. Each sublist has the following components:

<code>quality</code>	quality threshold used in <code>read_sample</code> .
<code>reference</code>	reference ID.
<code>centers</code>	a data frame with the centroids coordinates.
<code>data</code>	a data frame with the fluorescence intensities.

Examples

```
library(dPCP)

#Find path of sample table and location of reference and input files
sampleTable <- system.file("extdata", "Template_sampleTable.csv",
                           package = "dPCP")

fileLoc <- system.file("extdata",package = "dPCP")

#Read sample table file
sample.table <- read_sampleTable(sampleTable, system = "bio-rad",
                                 file.location = fileLoc)

#Read reference files
ref <- read_reference(sample.table, system = "bio-rad",
                     file.location = fileLoc)

#Read samples files
samp <- read_sample(sample.table, system = "bio-rad",
                   file.location = fileLoc)

#Reference DBSCAN clustering
dbref <- reference_dbscan(ref, sample.table, save.template = FALSE)

#Predict position of clusters centroid from reference DBSCAN results
cent <- centers_data(samp, sample.table,dbref)

plot(cent, sample = "all")
```

Description

This function carries out the c-means cluster analysis, using the centroids position as initial values for cluster centers.

Usage

```
cmeans_clus(centers.data)

## S3 method for class 'cmeans_clus'
plot(x, ..., sample = "all")
```

Arguments

centers.data	an object of class <code>centers_data</code> , inherited from <code>centers_data</code> .
x	an object of class <code>cmeans_clus</code>
...	Arguments to be passed to methods
sample	'all' to show all samples, or a numeric vector indicating the row number of samples in the sample table.

Value

An object of class `cmeans_clus` containing a sublist for each sample. Each sublist has the following components:

quality	quality threshold used in <code>read_sample</code> .
reference	reference ID.
centers	a data frame with the centroids coordinates.
data	a data frame with the fluorescence intensities and clusters name.
membership	a matrix with the membership values of the data elements to the clusters. See also <code>cmeans</code>

Examples

```
library(dPCP)

#Find path of sample table and location of reference and input files
sampleTable <- system.file("extdata", "Template_sampleTable.csv",
                           package = "dPCP")

fileLoc <- system.file("extdata", package = "dPCP")

#Read sample table file
sample.table <- read_sampleTable(sampleTable, system = "bio-rad",
                                file.location = fileLoc)

#Read reference files
ref <- read_reference(sample.table, system = "bio-rad",
                     file.location = fileLoc)

#Read samples files
samp <- read_sample(sample.table, system = "bio-rad",
                   file.location = fileLoc)
```

```

#Reference DBSCAN clustering
dbref <- reference_dbscan(ref, sample.table, save.template = FALSE)

#Predict position of clusters centroid from reference DBSCAN results
cent <- centers_data(samp, sample.table,dbref)

#Fuzzy c-means clustering
cmclus <- cmeans_clus(cent)

plot(cmclus, sample = "all")

```

dbscan_combination *Test eps and minPts combinations for DBSCAN analysis*

Description

This function tests all combinations of eps and minPts for DBSCAN analysis of reference samples indicated in refID. The results are represented in scatterplots exported to a pdf file.

Usage

```

dbscan_combination(
  refID,
  system = NULL,
  file.location = ".",
  reference.quality = 0.5,
  eps = c(120, 150, 180, 200),
  minPts = c(20, 50, 80, 100)
)

```

Arguments

refID	a string or a character vector of chipID (Thermo Fisher) or the complete file name with the extension (Bio-Rad) of reference sample(s) to be analysed.
system	character. The name of digital PCR system used to generate the data. It must be either Thermo Fisher or Bio-Rad. Abbreviations are also accepted.
file.location	character. Full path name to reference and sample files location. The default corresponds to the working directory, (getwd). Tilde expansion (see (path.expand)) is performed.
reference.quality	numeric. Between 0 and 1. Quality threshold to subset the data (just for Thermo Fisher). If different thresholds have to be applied to various reference samples, a vector of the same length of refID has to be provided.
eps	a numeric vector of values to be tested. Maximum distance between elements within a cluster in a DBSCAN analysis. See also dbscan .
minPts	a numeric vector of values to be tested. Number of minimum elements to assemble a cluster in a DBSCAN analysis. See also dbscan .

Value

A pdf file containing the scatterplots of DBSCAN analysis performed with all combinations of eps and minPts. Each reference generates a different pdf file.

Examples

```
library(dPCP)

#Find path of sample table and location of reference and input files
sampleTable <- system.file("extdata", "Template_sampleTable.csv",
                           package = "dPCP")

fileLoc <- system.file("extdata", package = "dPCP")

dbscan_combination("dilution20200313_B01_Amplitude.csv",
                  file.location = fileLoc, system = "bio-rad",
                  eps = c(150, 160, 180, 190), minPts = c(80, 100, 120))
```

dPCP

Automated analysis of digital PCR data

Description

This function carries out the automated clustering of digital PCR data.

Usage

```
dPCP(
  file,
  system = NULL,
  file.location = ".",
  reference.quality = 0.5,
  sample.quality = 0.5,
  eps = 200,
  minPts = 50,
  save.template = FALSE,
  rain = TRUE
)

## S3 method for class 'dPCP'
plot(x, ..., sample = "all", reference = "all", type = "dPCP")
```

Arguments

<code>file</code>	character. The name or the path of csv file to be read. If it does not contain an absolute path, the file name is relative to the current working directory, (getwd).
<code>system</code>	character. The name of digital PCR system used to generate the data. It must be either Thermo Fisher or Bio-Rad. Abbreviations are also accepted.
<code>file.location</code>	character. Full path name to reference and sample files location. The default corresponds to the working directory, (getwd). Tilde expansion (see (path.expand)) is performed.
<code>reference.quality</code>	numeric. Between 0 and 1. Quality threshold to subset the data. If different thresholds have to be applied to various reference samples, a vector of the same length of number of reference samples has to be provided. Used only when the system is Thermo Fisher.
<code>sample.quality</code>	numeric. Between 0 and 1. Quality threshold to subset data. If different thresholds have to be applied to various samples, a vector of the same length of number of samples has to be provided. Used only when the system is Thermo Fisher.
<code>eps</code>	numeric. Input parameter for the DBSCAN algorithm. It represents the maximum distance between the elements within a cluster. See also dbscan . If different values have to be applied to various reference samples, a vector of the same length of number of reference samples has to be provided.
<code>minPts</code>	numeric. Input parameter for the DBSCAN algorithm. It represents the number of minimum elements to assemble a cluster. See also dbscan . If different values have to be applied to various reference samples, a vector of the same length of number of reference samples has to be provided.
<code>save.template</code>	logical. If TRUE a template of DBSCAN analysis of reference samples is saved. When system is Thermo Fisher, <code>save.template</code> can be also a character vector indicating the chipID.
<code>rain</code>	logical. If TRUE the rain analysis is carried out.
<code>x</code>	an object of class dPCP
<code>...</code>	Arguments to be passed to methods
<code>sample</code>	'all' to show all samples, or a numeric vector indicating the row number of samples in the sample table.
<code>reference</code>	'all' to show all reference samples, or a character vector with chip ID (Thermo Fisher) or the file name (Bio-rad) of reference samples to be showed.
<code>type</code>	string. Type of plot to be showed. Available plots: 'reference dbscan', 'centers', 'cmeans', 'rain', 'dPCP'.

Value

An object of class dPCP containing the following components:

<code>referenceDB</code>	an object of class <code>reference_dbscan</code> .
<code>samples</code>	a list of samples. Each sample sublist contains the information about the cluster analysis.
<code>results</code>	an object of class <code>replicates_quant</code> .

Examples

```
library(dPCP)

#Find path of sample table and location of reference and input files
sampleTable <- system.file("extdata", "Template_sampleTable.csv",
                           package = "dPCP")

fileLoc <- system.file("extdata", package = "dPCP")

#dPCP analysis
results <- dPCP(sampleTable, system = "bio-rad", file.location = fileLoc,
                eps = 200, minPts = 50, save.template = FALSE, rain = TRUE)

plot(results, sample = 1, type = "dPCP")
```

export_csv

Export dPCP analysis results to a csv file

Description

This function exports dPCP analysis results to a csv file.

Usage

```
export_csv(data, filename)
```

Arguments

data an object of class dPCP, target_quant or replicates_quant.
filename character. File name (no extension) for csv and pdf files to create on disk.

Value

A csv file with the information and results of dPCP analysis.

Examples

```
library(dPCP)

#Find path of sample table and location of reference and input files
sampleTable <- system.file("extdata", "Template_sampleTable.csv",
                           package = "dPCP")

fileLoc <- system.file("extdata", package = "dPCP")

#dPCP analysis
```



```
results <- dPCP(sampleTable, system = "bio-rad", file.location = fileLoc,  
               eps = 200, minPts = 50, save.template = FALSE,  
               rain = TRUE)  
  
export_csv(results, filename = "dPCRproject_1")
```

manual_correction *Manual correction of dPCP cluster analysis*

Description

This function builds an interactive app to manually correct the dPCP cluster analysis.

Usage

```
manual_correction(data, filename, save.plot = FALSE, format = "png", dpi = 300)
```

Arguments

data	an object of class dPCP, inherited from dPCP .
filename	character. File name (no extension) for csv and pdf files to create on disk.
save.plot	logical. If TRUE the plots are exported to a file.
format	a string indicating the file format for the export. Available formats: 'eps', 'ps', 'tex', 'pdf', 'jpeg', 'tiff', 'png', 'bmp', 'svg', 'wmf'.
dpi	numeric. Image resolution.

Value

A Shiny session.

Examples

```
library(dPCP)  
  
#Find path of sample table and location of reference and input files  
sampleTable <- system.file("extdata", "Template_sampleTable.csv",  
                           package = "dPCP")  
  
fileLoc <- system.file("extdata", package = "dPCP")  
  
#dPCP analysis  
results <- dPCP(sampleTable, system = "bio-rad", file.location = fileLoc,  
               eps = 200, minPts = 50, save.template = FALSE,  
               rain = TRUE)  
  
manual_correction(results, filename = "manual_dPCR", save.plot = FALSE)
```



```
#Read reference files
ref <- read_reference(sample.table, system = "bio-rad",
                     file.location = fileLoc)

#Read samples files
samp <- read_sample(sample.table, system = "bio-rad",
                   file.location = fileLoc)

#Reference DBSCAN clustering
dbref <- reference_dbscan(ref, sample.table, save.template = FALSE)

#Predict position of clusters centroid from reference DBSCAN results
cent <- centers_data(samp, sample.table,dbref)

#Fuzzy c-means clustering
cmclus <- cmeans_clus(cent)

#Rain classification.
rainclus <- rain_reclus(cmclus)

plot(rainclus, sample = "all")
```

read_reference	<i>Read reference files</i>
----------------	-----------------------------

Description

This function reads the results files of reference samples listed in the sample table. Fluorescence intensity and quality value (just for Thermo Fisher) are collected. If a [reference_dbscan](#) template file with the same input parameters (reference ID, eps, minPts) is available, fluorescence data, quality value and dbscan analysis results are retrieved from the template file.

Usage

```
read_reference(
  sample.table,
  system = NULL,
  file.location = ".",
  reference.quality = 0.5,
  eps = NULL,
  minPts = NULL
)
```

Arguments

`sample.table` object of class `sample_table`, inherited from [read_sampleTable](#).

system	character. The name of digital PCR system used to generate the data. It must be either Thermo Fisher or Bio-Rad. Abbreviations are also accepted.
file.location	character. Full path name to reference and sample files location. The default corresponds to the working directory, (<code>getwd</code>). Tilde expansion (see (<code>path.expand</code>)) is performed.
reference.quality	numeric. Between 0 and 1. Quality threshold to subset the data. If different thresholds have to be applied to various reference samples, a vector of the same length of number of reference samples has to be provided. Used only when the system is Thermo Fisher.
eps, minPts	numeric. Input parameters for the DBSCAN algorithm. If they match the parameters of <code>reference_dbscan</code> template file, the data are retrieved from the template.

Value

An object of class `read_reference` containing a sublist for each reference. Each sublist has the following components:

quality	value of the <code>reference.quality</code> parameter.
data	a matrix with the fluorescence intensities and quality values.
dbscan	an object of class <code>dbscan_fast</code> , inherited from <code>dbscan</code> . This component is available only if a <code>reference_dbscan</code> template file is used to retrieve the data.

Examples

```
library(dPCP)

#Find path of sample table and location of reference and input files
sampleTable <- system.file("extdata", "Template_sampleTable.csv",
                           package = "dPCP")

fileLoc <- system.file("extdata", package = "dPCP")

#Read sample table file
sample.table <- read_sampleTable(sampleTable, system = "bio-rad",
                                 file.location = fileLoc)

#Read reference files
ref <- read_reference(sample.table, system = "bio-rad",
                     file.location = fileLoc)
```

read_sample

Read sample files

Description

This function reads the results files of samples listed in the sample table. Fluoresce intensity and quality value (just for Thermo Fisher) are collected.

Usage

```
read_sample(
  sample.table,
  system = NULL,
  file.location = ".",
  sample.quality = 0.5
)
```

Arguments

`sample.table` object of class `sample_table`, inherited from `read_sampleTable`.

`system` character. The name of digital PCR system used to generate the data. It must be either Thermo Fisher or Bio-Rad. Abbreviations are also accepted.

`file.location` character. Full path name to reference and sample files location. The default corresponds to the working directory, (`getwd`). Tilde expansion (see (`path.expand`)) is performed.

`sample.quality` numeric. Between 0 and 1. Quality threshold to subset data. If different thresholds have to be applied to various samples, a vector of the same length of number of samples has to be provided. Used only when the system is Thermo Fisher.

Value

An object of class `read_sample` containing a sublist for each sample. Each sublist has the following components:

`quality` value of the `sample.quality` parameter.

`data` a matrix with the fluorescence intensities and quality values.

Examples

```
library(dPCP)

#Find path of sample table and location of reference and input files
sampleTable <- system.file("extdata", "Template_sampleTable.csv",
  package = "dPCP")

fileLoc <- system.file("extdata", package = "dPCP")

#Read sample table file
sample.table <- read_sampleTable(sampleTable, system = "bio-rad",
  file.location = fileLoc)

#Read reference files
ref <- read_reference(sample.table, system = "bio-rad",
  file.location = fileLoc)

#Read samples files
samp <- read_sample(sample.table, system = "bio-rad",
  file.location = fileLoc)
```

reference_dbscan	<i>Find the empty partitions and single target clusters in the reference sample</i>
------------------	---

Description

This function computes a DBSCAN analysis to identify single target clusters in the reference samples listed in the sample table. If a `reference_dbscan` template file with the same input parameters (reference ID, eps, minPts) is available, data are retrieved from the template file.

Usage

```
reference_dbscan(
  reference.subquality,
  sample.table,
  eps = 200,
  minPts = 50,
  save.template = FALSE
)

## S3 method for class 'reference_dbscan'
plot(x, ..., reference = "all")
```

Arguments

reference.subquality	an object of class <code>read_reference</code> , inherited from <code>read_reference</code> .
sample.table	object of class <code>sample_table</code> , inherited from <code>read_sampleTable</code> .
eps	numeric. Input parameters for the DBSCAN algorithm. If they match the parameters of <code>reference_dbscan</code> template file, the data are retrieved from the template.
minPts	numeric. Input parameters for the DBSCAN algorithm. If they match the parameters of <code>reference_dbscan</code> template file, the data are retrieved from the template.
save.template	logical. If TRUE a template of DBSCAN analysis of reference samples is saved. When system is Thermo Fisher, <code>save.template</code> can be also a character vector indicating the chipID.
x	an object of class <code>reference_dbscan</code>
...	Arguments to be passed to methods
reference	'all' to show all reference samples, or a character vector with chip ID (Thermo Fisher) or the file name (Bio-rad) of reference samples to be showed.

Value

An object of class `reference_dbscan` containing a sublist for each reference. Each sublist has the following components:

<code>quality</code>	quality threshold used in <code>read_reference</code> .
<code>data</code>	a matrix with the fluorescence intensities and quality values.
<code>dbscan</code>	an object of class <code>dbscan_fast</code> , inherited from <code>dbscan</code> .

Examples

```
library(dPCP)

#Find path of sample table and location of reference and input files
sampleTable <- system.file("extdata", "Template_sampleTable.csv",
                           package = "dPCP")

fileLoc <- system.file("extdata", package = "dPCP")

#Read sample table file
sample.table <- read_sampleTable(sampleTable, system = "bio-rad",
                                 file.location = fileLoc)

#Read reference files
ref <- read_reference(sample.table, system = "bio-rad",
                     file.location = fileLoc)

#Read samples files
samp <- read_sample(sample.table, system = "bio-rad",
                   file.location = fileLoc)

#Reference DBSCAN clustering
dbref <- reference_dbscan(ref, sample.table, save.template = FALSE)

plot(dbref, reference = "all")
```

 replicates_quant

Calculation of targets concentration, pooling the sample replicates

Description

This function calculates the concentration of the targets, combining the results of the replicates of each sample.

Usage

```
replicates_quant(raw.results, sample.table)
```


Arguments

`raw.results` an object of class `target_quant`, inherited from `target_quant`.
`sample.table` object of class `sample_table`, inherited from `read_sampleTable`.

Value

An object of class `replicates_quant` containing a sublist for every sample. Each sublist has the following components:

`quality` quality threshold used in `read_sample`.
`reference` reference ID.
`raw.results` a data frame with the results of quantification.
`replicates.results`
a data frame with the results of quantification of pooled replicates.

Examples

```
library(dPCP)

#Find path of sample table and location of reference and input files
sampleTable <- system.file("extdata", "Template_sampleTable.csv",
                           package = "dPCP")

fileLoc <- system.file("extdata", package = "dPCP")

#Read sample table file
sample.table <- read_sampleTable(sampleTable, system = "bio-rad",
                                file.location = fileLoc)

#Read reference files
ref <- read_reference(sample.table, system = "bio-rad",
                     file.location = fileLoc)

#Read samples files
samp <- read_sample(sample.table, system = "bio-rad",
                   file.location = fileLoc)

#Reference DBSCAN clustering
dbref <- reference_dbscan(ref, sample.table, save.template = FALSE)

#Predict position of clusters centroid from reference DBSCAN results
cent <- centers_data(samp, sample.table,dbref)

#Fuzzy c-means clustering
cmclus <- cmeans_clus(cent)

#Rain classification.
rainclus <- rain_reclus(cmclus)
```

```
#Quantification
quantcm <- target_quant(cmclus, sample.table)
quant <- target_quant(rainclus, sample.table)

#Replicates pooling
rep.quant <- replicates_quant(quant, sample.table)
```

report_dPCP

Export dPCP analysis results to a pdf report

Description

This function generates a pdf report of the dPCP analysis.

Usage

```
report_dPCP(data, filename, sample = "all")
```

Arguments

data	an object of class dPCP, inherited from dPCP .
filename	character. File name (no extension) for csv and pdf files to create on disk.
sample	'all' to show all samples, or a numeric vector indicating the row number of samples in the sample table.

Value

A pdf file with the information and results of the dPCP analysis.

Examples

```
library(dPCP)

#Find path of sample table and location of reference and input files
sampleTable <- system.file("extdata", "Template_sampleTable.csv",
                           package = "dPCP")

fileLoc <- system.file("extdata", package = "dPCP")

#dPCP analysis
results <- dPCP(sampleTable, system = "bio-rad", file.location = fileLoc,
               eps = 200, minPts = 50, save.template = FALSE,
               rain = TRUE)

report_dPCP(results, filename = "dPCRproject_1")
```

target_quant	<i>Calculation of targets concentration.</i>
--------------	--

Description

This function calculates the concentration of the targets according to the Poisson distribution.

Usage

```
target_quant(data.cluster, sample.table)
```

Arguments

`data.cluster` an object of class `rain_reclus` or `cmeans_clus`.
`sample.table` object of class `sample_table`, inherited from [read_sampleTable](#).

Value

An object of class `target_quant` containing a sublist for each sample. Each sublist has the following components:

`quality` quality threshold used in [read_sample](#).
`reference` reference ID.
`raw results` a data frame with the results of the quantification.

Examples

```
library(dPCP)

#Find path of sample table and location of reference and input files
sampleTable <- system.file("extdata", "Template_sampleTable.csv",
                           package = "dPCP")

fileLoc <- system.file("extdata", package = "dPCP")

#Read sample table file
sample.table <- read_sampleTable(sampleTable, system = "bio-rad",
                                 file.location = fileLoc)

#Read reference files
ref <- read_reference(sample.table, system = "bio-rad",
                      file.location = fileLoc)

#Read samples files
samp <- read_sample(sample.table, system = "bio-rad",
                    file.location = fileLoc)

#Reference DBSCAN clustering
```

```
dbref <- reference_dbscan(ref, sample.table, save.template = FALSE)

#Predict position of clusters centroid from reference DBSCAN results
cent <- centers_data(samp, sample.table,dbref)

#Fuzzy c-means clustering
cmclus <- cmeans_clus(cent)

#Rain classification.
rainclus <- rain_reclus(cmclus)

#Quantification
quantcm <- target_quant(cmclus, sample.table)
quant <- target_quant(rainclus, sample.table)
```

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