Package 'ncvreg'

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Title Regularization Paths for SCAD and MCP Penalized Regression Models

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Description Fits regularization paths for linear regression, GLM, and Cox regression models using lasso or nonconvex penalties, in particular the minimax concave penalty (MCP) and smoothly clipped absolute deviation (SCAD) penalty, with options for additional L2 penalties (the ``elastic net" idea). Utilities for carrying out cross-validation as well as post-fitting visualization, summarization, inference, and prediction are also provided. For more information, see Breheny and Huang (2011) <doi:10.1214/10-AOAS388> or visit the nevreg homepage https://pbreheny.github.io/nevreg/.

BugReports https://github.com/pbreheny/ncvreg/issues

License GPL-3

URL https://pbreheny.github.io/ncvreg/,
 https://github.com/pbreheny/ncvreg

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2 ncvreg-package

R topics documented:

ncvreg-package		Regularization paths for SCAD- and MCP-penalized regression models											d-											
Index																								38
	summary.ncvreg				•		•	• •	•		•	٠	 ٠	•	 •	 •	•	•	•	 •	•	•		3.
	summary.cv.ncvreg																							
	std																							
	Prostate																							
	predict.ncvsurv																							
	predict.ncvreg																							
	plot.ncvsurv.func .																							
	plot.ncvreg																							
	plot.mfdr																							
	plot.cv.ncvreg																							
	permres																							
	perm.ncvreg																							
	ncvsurv																							
	ncvreg																							
	ncvfit																							12
	mfdr																							10
	Lung																							9
	local_mfdr																							
	Heart																							•
	fir																							
	cv.ncvreg																							
	AUC.cv.ncvsurv																							
	ncvreg-package																							1

Description

Efficient algorithms for fitting regularization paths for a variety of regression models (linear, logistic, Poisson, survival) penalized by MCP or SCAD, with optional additional L2 penalty.

Details

Accepts a design matrix X and vector of responses y, produces the regularization path over a grid of values for the tuning parameter lambda. Also provides methods for plotting, cross-validation-based inference, and for determining locally convex regions of the coefficients paths.

See the "Getting started" vignette for a brief overview of how the package works.

Visit the nevreg website for more details.

Author(s)

Patrick Breheny patrick-breheny@uiowa.edu>

AUC.cv.ncvsurv 3

References

Breheny P and Huang J. (2011) Coordinate descentalgorithms for nonconvex penalized regression, with applications to biological feature selection. *Annals of Applied Statistics*, **5**: 232-253. doi: 10.1214/10AOAS388

Examples

```
## Not run:
vignette("getting-started", package="ncvreg")
## End(Not run)
```

AUC.cv.ncvsurv

Calculates AUC for cv.ncvsurv objects

Description

Calculates the cross-validated AUC (concordance) from a "cv.ncvsurv" object.

Usage

```
## S3 method for class 'cv.ncvsurv'
AUC(obj, ...)
```

Arguments

obj A cv.ncvsurv object. You must run cv.ncvsurv with the option returnY=TRUE

in order for AUC to work.

... For S3 method compatibility?

Details

The area under the curve (AUC), or equivalently, the concordance statistic (C), is calculated according to the procedure outlined in the reference below. This calls the survConcordance function in the survival package, except the cross-validated linear predictors are used to guard against overfitting. Thus, the values returned by AUC.cv.ncvsurv will be lower than those you would obtain with survConcordance if you fit the full (unpenalized) model.

Author(s)

Patrick Breheny, Brandon Butcher, and Lawrence Hunsicker

References

van Houwelingen H, Putter H (2011). Dynamic Prediction in Clinical Survival Analysis. CRC Press.

4 cv.ncvreg

See Also

```
cv.ncvsurv, survConcordance
```

Examples

cv.ncvreg

Cross-validation for nevreg/nevsurv

Description

Performs k-fold cross validation for MCP- or SCAD-penalized regression models over a grid of values for the regularization parameter lambda.

Usage

```
cv.ncvreg(X, y, ..., cluster, nfolds=10, seed, fold, returnY=FALSE,
trace=FALSE)
cv.ncvsurv(X, y, ..., cluster, nfolds=10, seed, fold, se=c('quick', 'bootstrap'), returnY=FALSE, trace=FALSE)
```

Arguments

seed

Χ	The design matrix, without an intercept, as in ncvreg/ncvsurv.
у	The response vector, as in nevreg/nevsurv.
	Additional arguments to ncvreg/ncvsurv.
cluster	cv.ncvreg and cv.ncvsurv can be run in parallel across a cluster using the parallel package. The cluster must be set up in advance using the makeCluster function from that pacakge. The cluster must then be passed to cv.ncvreg/cv.ncvsurv (see example).
nfolds	The number of cross-validation folds. Default is 10.
fold	Which fold each observation belongs to. By default the observations are randomly assigned

You may set the seed of the random number generator in order to obtain repro-

ducible results.

cv.ncvreg 5

returnY Should cv.ncvreg/cv.ncvsurv return the linear predictors from the cross-validation

folds? Default is FALSE; if TRUE, this will return a matrix in which the element for row i, column j is the fitted value for observation i from the fold in which observation i was excluded from the fit, at the jth value of lambda. NOTE: For cv.ncvsurv, the rows of Y are ordered by time on study, and therefore will not

correspond to the original order of observations pased to cv.ncvsurv.

If set to TRUE, inform the user of progress by announcing the beginning of each

CV fold. Default is FALSE.

se For cv.ncvsurv, the method by which the cross-valiation standard error (CVSE)

is calculated. The 'quick' approach is based on a rough approximation, but can be calculated more or less instantly. The 'bootstrap' approach is more accurate,

but requires additional computing time.

Details

trace

The function calls nevreg/nevsurv nfolds times, each time leaving out 1/nfolds of the data. The cross-validation error is based on the deviance; see here for more details.

For family="binomial" models, the cross-validation fold assignments are balanced across the 0/1 outcomes, so that each fold has the same proportion of 0/1 outcomes (or as close to the same proportion as it is possible to achieve if cases do not divide evenly).

For Cox models, cv.ncvsurv uses the approach of calculating the full Cox partial likelihood using the cross-validated set of linear predictors. Other approaches to cross-validation for the Cox regression model have been proposed in the literature; the strengths and weaknesses of the various methods for penalized regression in the Cox model are the subject of current research. A simple approximation to the standard error is provided, although an option to bootstrap the standard error (se='bootstrap') is also available.

Value

An object with S3 class cv.ncvreg/cv.ncvsurv containing:

cve The error for each value of lambda, averaged across the cross-validation folds.

cvse The estimated standard error associated with each value of for cve.

fold The fold assignments for cross-validation for each observation; note that for cv.ncvsurv, these are in terms of the ordered observations, not the original observations.

lambda The sequence of regularization parameter values along which the cross-validation error was calculated.

fit The fitted nevreg/nevsury object for the whole data.

min The index of lambda corresponding to lambda.min.

lambda.min The value of lambda with the minimum cross-validation error.

null.dev The deviance for the intercept-only model. If you have supplied your own lambda sequence, this quantity may not be meaningful.

Bias The estimated bias of the minimum cross-validation error, as in Tibshirani RJ and Tibshirani R (2009), "A Bias Correction for the Minimum Error Rate in Cross-Validation", Ann. Appl. Stat. 3:822-829.

pe If family="binomial", the cross-validation prediction error for each value of lambda.

Y If returnY=TRUE, the matrix of cross-validated fitted values (see above).

6 fir

Author(s)

Patrick Breheny; Grant Brown helped with the parallelization support

References

Breheny P and Huang J. (2011) Coordinate descentalgorithms for nonconvex penalized regression, with applications to biological feature selection. *Annals of Applied Statistics*, **5**: 232-253. doi: 10.1214/10AOAS388

See Also

```
ncvreg, plot.cv.ncvreg, summary.cv.ncvreg
```

Examples

```
data(Prostate)
cvfit <- cv.ncvreg(Prostate$X, Prostate$y)</pre>
plot(cvfit)
summary(cvfit)
fit <- cvfit$fit</pre>
plot(fit)
beta <- fit$beta[,cvfit$min]</pre>
## requires loading the parallel package
## Not run:
library(parallel)
X <- Prostate$X
y <- Prostate$y
cl <- makeCluster(4)</pre>
cvfit <- cv.ncvreg(X, y, cluster=cl, nfolds=length(y))</pre>
## End(Not run)
# Survival
data(Lung)
X <- Lung$X
y <- Lung$y
cvfit <- cv.ncvsurv(X, y)</pre>
summary(cvfit)
plot(cvfit)
plot(cvfit, type="rsq")
```

fir

Marginal false discovery rates

Description

Estimates the marginal false discovery rate (mFDR) of a penalized regression model.

Heart 7

Usage

```
fir(fit, ...)
```

Arguments

fit An nowneg or nowsury object.
... Arguments to pass to mfdr.

Details

This function has been renamed and is currently deprecated. Use mfdr instead.

Heart

Risk factors associated with heart disease

Description

Data from a subset of the Coronary Risk-Factor Study baseline survey, carried out in rural South Africa.

- X: A design matrix with 462 observations (rows) and 9 predictor variables (columns). The columns are:
 - sbp: Systolic blood pressure
 - tobacco: Cumulative tobacco consumption, in kg
 - ldl: Low-density lipoprotein cholesterol
 - adiposity: Adipose tissue concentration
 - famhist: Family history of heart disease (1=Present, 0=Absent)
 - typea: Score on test designed to measure type-A behavior
 - obesity: Obesity
 - alcohol: Current consumption of alcohol
 - age: Age of subject
- y: Coronary heart disease at baseline; 1=Yes 0=No

Usage

```
data(Heart)
```

Source

https://web.stanford.edu/~hastie/ElemStatLearn/

References

- Hastie T, Tibshirani R, and Friedman J. (2001). The Elements of Statistical Learning. Springer.
- Rousseauw J, et al. (1983). Coronary risk factor screening in three rural communities. *South African Medical Journal*, **64**, 430-436.

8 local_mfdr

local_mfdr Estimate local mFDR for all features	
---	--

Description

local_mfdr() is called by summary.ncvreg(), which typically offers a more convenient interface to users. If, however, you are working with local mfdrs programmatically rather than interactively, you probably want to use local_mfdr(), which skips the sorting, filtering, and print formatting of summary.ncvreg().

Usage

```
local_mfdr(fit, lambda, X = NULL, y = NULL, method = c("ashr", "kernel"), ...)
```

Arguments

fit	A fitted novreg or novsurv object.
lambda	The value of lambda at which inference should be carried out.
X, y	The design matrix and response used to fit the model; in most cases, it is not necessary to provide X and y as they are returned by ncvreg, but see the returnX argument in ncvreg().
method	What method should be used to calculate the local fdr? Options are ashr (which tends to be more accurate) and kernel (which requires no additional packages). The default is to use ashr if the package is installed.
	Additional arguments to ash() if using method='ashr'.

Value

If all features are penalized, then the object returns a data frame with one row per feature and four columns:

- Estimate: The coefficient estimate from the penalized regression fit
- z: A test statistic that approximately follows a standard normal distribution under the null hypothesis that the feature is marginally independent of the outcome
- mfdr: The estimated marginal local false discovery rate
- Selected: Features with nonzero coefficient estimates are given an asterisk

If some features are penalized and others are not, then a list is returned with two elements: pen.vars, which consists of the data frame described above, and unpen.vars, a data frame with four columns: Estimate, SE, Statistic, and p.value. The standard errors and p-values are based on a classical lm/glm/coxph model using the effect of the penalized features as an offset.

See Also

```
summary.ncvreg()
```

Lung 9

Examples

```
# Linear regression
data(Prostate)
fit <- ncvreg(Prostate$X, Prostate$y)</pre>
local_mfdr(fit, 0.1)
fit <- ncvreg(Prostate$X, Prostate$y, penalty.factor=rep(0:1, each=4))</pre>
local_mfdr(fit, 0.1)
# Logistic regression
data(Heart)
X <- Heart$X
y <- Heart$y
fit <- ncvreg(X, y, family='binomial')</pre>
local_mfdr(fit, 0.1)
# Cox regression
data(Lung)
X <- Lung$X
y <- Lung$y
fit <- ncvsurv(X, y)</pre>
local_mfdr(fit, 0.1)
```

Lung

VA lung cancer data set

Description

Data from a randomised trial of two treatment regimens for lung cancer. This is a standard survival analysis data set from the classic textbook by Kalbfleisch and Prentice.

- X: A design matrix with 137 observations (rows) and 9 predictor variables (columns). The columns are:
 - trt: Treatment indicator (1=control group, 2=treatment group)
 - karno: Karnofsky performance score (0=bad, 100=good)
 - diagtime: Time from diagnosis to randomization (months)
 - age: Age (years)
 - prior: Prior therapy (0=no, 1=yes)
 - squamous: Indicator for whether the cancer type is squamous cell carcinoma (0=no, 1=yes)
 - small: Indicator for whether the cancer type is small cell lung cancer (0=no, 1=yes)
 - adeno: Indicator for whether the cancer type is adenocarcinoma (0=no, 1=yes)
 - large: Indicator for whether the cancer type is large cell carcinoma (0=no, 1=yes)
- y: A two column matrix (Surv object) containing the follow-up time (in days) and an indicator variable for whether the patient died while on the study or not.

10 mfdr

Usage

```
data(Lung)
```

Format

A list containing the design matrix X and response matrix y

Source

```
https://cran.r-project.org/package=survival
```

References

• Kalbfleisch D and Prentice RL (1980), *The Statistical Analysis of Failure Time Data*. Wiley, New York.

See Also

ncvsurv

mfdr

Marginal false discovery rates

Description

Estimates the marginal false discovery rate (mFDR) of a penalized regression model.

Usage

```
mfdr(fit, X)
```

Arguments

fit An novreg or novsury object.

Χ

The model matrix corresponding to fit. This is not necessary for linear regression, but in logistic and Cox regression, the mFDR depends on X. It is not necessary to supply X if it is already contained in fit; i.e., if ncvreg/ncvsurv was run with returnX=TRUE.

Details

The function estimates the marginal false discovery rate (mFDR) for a penalized regression model. The estimate tends to be accurate in most settings, but will be slightly conservative if predictors are highly correlated. For an alternative way of estimating the mFDR, typically more accurate in highly correlated cases, see perm.ncvreg.

mfdr 11

Value

An object with S3 class mfdr inheriting from data. frame and containing:

EF The number of variables selected at each value of lambda, averaged over the

permutation fits.

S The actual number of selected variables for the non-permuted data.

mFDR The estimated marginal false discovery rate (EF/S).

Author(s)

Patrick Breheny and Ryan Miller

See Also

```
ncvreg, ncvsurv, plot.mfdr, perm.ncvreg
```

Examples

```
# Linear regression -----
data(Prostate)
fit <- ncvreg(Prostate$X, Prostate$y)</pre>
obj <- mfdr(fit)</pre>
obj[1:10,]
# Comparison with perm.ncvreg
op <- par(mfrow=c(2,2))</pre>
plot(obj)
plot(obj, type="EF")
pmfit <- perm.ncvreg(Prostate$X, Prostate$y)</pre>
plot(pmfit)
plot(pmfit, type="EF")
par(op)
# Logistic regression -----
data(Heart)
fit <- ncvreg(Heart$X, Heart$y, family="binomial")</pre>
obj <- mfdr(fit)</pre>
head(obj)
op <- par(mfrow=c(1,2))
plot(obj)
plot(obj, type="EF")
par(op)
# Cox regression -----
data(Lung)
fit <- ncvsurv(Lung$X, Lung$y)</pre>
obj <- mfdr(fit)</pre>
head(obj)
op <- par(mfrow=c(1,2))
```

12 ncvfit

```
plot(obj)
plot(obj, type="EF")
par(op)
```

ncvfit

Direct interface for nonconvex penalized regression (non-pathwise)

Description

This function is intended for users who know exactly what they're doing and want complete control over the fitting process: no standardization is applied, no intercept is included, no path is fit. All of these things are best practices for data analysis, so if you are choosing not to do them, you are on your own – there is no guarantee that your results will be meaningful. Some things in particular that you should pay attention to:

- If your model has an intercept, it is up to you to (un)penalize it properly, typically by settings its corresponding element of penalty. factor to zero.
- You should provide initial values for the coefficients; in nonconvex optimization, initial values are very important in determining which local solution an algorithm converges to.

Usage

```
ncvfit(
    X,
    y,
    init = rep(0, ncol(X)),
    r,
    xtx,
    penalty = c("MCP", "SCAD", "lasso"),
    gamma = switch(penalty, SCAD = 3.7, 3),
    alpha = 1,
    lambda,
    eps = 1e-05,
    max.iter = 1000,
    penalty.factor = rep(1, ncol(X)),
    warn = TRUE
)
```

Arguments

X	Design matrix; no intercept will be added, no standardization will occur (n x p matrix)
у	Response vector (length n vector)
init	Initial values for beta. Default: zero (length p vector)
r	Residuals corresponding to init; these will be calculated if not supplied, but if they have already been calculated elsewhere, it is more efficient to pass them as an argument. WARNING: If you supply an incorrect value of r, the solution

will be incorrect. (length n vector)

ncvfit 13

xtx	X scales: the jth element should equal $crossprod(X[,j])/n$. These will be calculated if not supplied, but if they have already been calculated elsewhere, it is more efficient to pass them as an argument. In particular, if X is standardized, one should pass $xtx = rep(1,p)$. WARNING: If you supply an incorrect value of xtx , the solution will be incorrect. (length p vector)
penalty	Penalty function to be applied, either "MCP" (default), "SCAD", or "lasso")
gamma	Tuning parameter of the MCP/SCAD penalty, as in $ncvreg()$; default is 3 for MCP and 3.7 for SCAD.
alpha	Tuning paramter controlling the ridge component of penalty, as in ncvreg(); default is 1 (meaning no ridge penalty)
lambda	Regularization parameter value at which to estimate beta; must be scalar – for pathwise optimization, see $ncvreg()$
eps	Convergence threshhold. The algorithm iterates until the RMSD for the change in linear predictors for each coefficient is less than eps. Default is 1e-4.
max.iter	Maximum number of allowed iterations; if this number is reached, algorithm will terminate prior to convergence. Default: 1000.
penalty.factor	Multiplicative factor for the penalty applied to each coefficient, as in ncvreg(). In particular, note that if you include an intercept, you probably want to set its entry to zero here.
warn	Return warning messages for failures to converge and model saturation? Default is TRUE.

Details

At the moment, this function only works for least-squares loss functions. Additional functionality for other loss functions (logistic, Cox) is in development.

Value

A list containing:

• beta: The estimated regression coefficients

 $\bullet\,$ iter: The number of iterations required to solve for 'beta

• loss: The loss (residual sum of squares) at convergence

• resid: The residuals at convergence

lambda: See abovepenalty: See abovegamma: See abovealpha: See above

• penalty.factor: See above

• n: Sample size

14 ncvreg

Examples

```
data(Prostate)
X <- cbind(1, Prostate$X)
y <- Prostate$y
fit <- ncvfit(X, y, lambda=0.1, penalty.factor=c(0, rep(1, ncol(X)-1)))
fit$beta
# Compare with:
coef(ncvreg(X, y), 0.1)
# The unstandardized version makes little sense here, as it fails to account
# for differences in the scales of the predictors.</pre>
```

ncvreg

Fit an MCP- or SCAD-penalized regression path

Description

Fit coefficients paths for MCP- or SCAD-penalized regression models over a grid of values for the regularization parameter lambda. Fits linear and logistic regression models, with option for an additional L2 penalty.

Usage

```
ncvreg(X, y, family=c("gaussian", "binomial", "poisson"),
penalty=c("MCP", "SCAD", "lasso"), gamma=switch(penalty, SCAD=3.7, 3),
alpha=1, lambda.min=ifelse(n>p,.001,.05), nlambda=100, lambda, eps=1e-4,
max.iter=10000, convex=TRUE, dfmax=p+1, penalty.factor=rep(1, ncol(X)),
warn=TRUE, returnX, ...)
```

Arguments

X	The design matrix, without an intercept. nevreg standardizes the data and includes an intercept by default.
у	The response vector.
family	Either "gaussian", "binomial", or "poisson", depending on the response.
penalty	The penalty to be applied to the model. Either "MCP" (the default), "SCAD", or "lasso".
gamma	The tuning parameter of the MCP/SCAD penalty (see details). Default is 3 for MCP and 3.7 for SCAD.
alpha	Tuning parameter for the Mnet estimator which controls the relative contributions from the MCP/SCAD penalty and the ridge, or L2 penalty. alpha=1 is equivalent to MCP/SCAD penalty, while alpha=0 would be equivalent to ridge regression. However, alpha=0 is not supported; alpha may be arbitrarily small, but not exactly 0.
lambda.min	The smallest value for lambda, as a fraction of lambda.max. Default is .001 if the number of observations is larger than the number of covariates and .05

otherwise.

ncvreg 15

nlambda	The number of lambda values. Default is 100.
lambda	A user-specified sequence of lambda values. By default, a sequence of values of length nlambda is computed, equally spaced on the log scale.
eps	Convergence threshhold. The algorithm iterates until the RMSD for the change in linear predictors for each coefficient is less than eps. Default is 1e-4.
max.iter	Maximum number of iterations (total across entire path). Default is 10000.
convex	Calculate index for which objective function ceases to be locally convex? Default is TRUE.
dfmax	Upper bound for the number of nonzero coefficients. Default is no upper bound. However, for large data sets, computational burden may be heavy for models with a large number of nonzero coefficients.
penalty.factor	A multiplicative factor for the penalty applied to each coefficient. If supplied, penalty.factor must be a numeric vector of length equal to the number of columns of X. The purpose of penalty.factor is to apply differential penalization if some coefficients are thought to be more likely than others to be in the model. In particular, penalty.factor can be 0, in which case the coefficient is always in the model without shrinkage.
warn	Return warning messages for failures to converge and model saturation? Default is TRUE.
returnX	Return the standardized design matrix along with the fit? By default, this option is turned on if X is under 100 MB, but turned off for larger matrices to preserve

Details

. . .

The sequence of models indexed by the regularization parameter lambda is fit using a coordinate descent algorithm. For logistic regression models, some care is taken to avoid model saturation; the algorithm may exit early in this setting. The objective function is defined to be

$$Q(\beta|X,y) = \frac{1}{n}L(\beta|X,y) + P_{\lambda}(\beta)$$

where the loss function L is the deviance (-2 times the log likelihood) for the specified outcome distribution (gaussian/binomial/poisson). See here for more details.

This algorithm is stable, very efficient, and generally converges quite rapidly to the solution. For GLMs, adaptive rescaling is used.

Value

An object with S3 class "novreg" containing:

Not used.

beta The fitted matrix of coefficients. The number of rows is equal to the number of coefficients, and the number of columns is equal to nlambda.

iter A vector of length nlambda containing the number of iterations until convergence at each value of lambda.

ncvreg

lambda The sequence of regularization parameter values in the path.

penalty Same as above.

family Same as above.

gamma Same as above.

alpha Same as above.

convex.min The last index for which the objective function is locally convex. The smallest value of lambda for which the objective function is convex is therefore lambda[convex.min], with corresponding coefficients beta[,convex.min].

loss A vector containing the deviance (i.e., the loss) at each value of lambda. Note that for gaussian models, the loss is simply the residual sum of squares.

penalty.factor Same as above.

n Sample size.

Additionally, if returnX=TRUE, the object will also contain

- **X** The standardized design matrix.
- y The response, centered if family='gaussian'.

Author(s)

Patrick Breheny

References

Breheny P and Huang J. (2011) Coordinate descentalgorithms for nonconvex penalized regression, with applications to biological feature selection. *Annals of Applied Statistics*, **5**: 232-253. doi: 10.1214/10AOAS388

See Also

```
plot.ncvreg, cv.ncvreg
```

Examples

ncvsurv 17

```
par(op)
op <- par(mfrow=c(2,2))</pre>
fit <- ncvreg(X, y)</pre>
plot(fit, main=expression(paste(alpha, "=",1)))
fit <- ncvreg(X, y, alpha=0.9)</pre>
plot(fit, main=expression(paste(alpha,"=",0.9)))
fit <- ncvreg(X, y, alpha=0.5)</pre>
plot(fit, main=expression(paste(alpha,"=",0.5)))
fit <- ncvreg(X, y, alpha=0.1)</pre>
plot(fit, main=expression(paste(alpha,"=",0.1)))
par(op)
op <- par(mfrow=c(2,2))</pre>
fit <- ncvreg(X, y)</pre>
plot(mfdr(fit))
                             # Independence approximation
plot(mfdr(fit), type="EF") # Independence approximation
perm.fit <- perm.ncvreg(X, y)</pre>
plot(perm.fit)
plot(perm.fit, type="EF")
par(op)
# Logistic regression ------
data(Heart)
X <- Heart$X
y <- Heart$y
op <- par(mfrow=c(2,2))</pre>
fit <- ncvreg(X, y, family="binomial")</pre>
plot(fit, main=expression(paste(gamma,"=",3)))
fit <- ncvreg(X, y, family="binomial", gamma=10)</pre>
plot(fit, main=expression(paste(gamma,"=",10)))
fit <- ncvreg(X, y, family="binomial", gamma=1.5)</pre>
plot(fit, main=expression(paste(gamma, "=", 1.5)))
fit <- ncvreg(X, y, family="binomial", penalty="SCAD")</pre>
plot(fit, main=expression(paste("SCAD, ",gamma,"=",3)))
par(op)
op <- par(mfrow=c(2,2))
fit <- ncvreg(X, y, family="binomial")</pre>
plot(fit, main=expression(paste(alpha,"=",1)))
fit <- ncvreg(X, y, family="binomial", alpha=0.9)</pre>
plot(fit, main=expression(paste(alpha,"=",0.9)))
fit <- ncvreg(X, y, family="binomial", alpha=0.5)</pre>
plot(fit, main=expression(paste(alpha,"=",0.5)))
fit <- ncvreg(X, y, family="binomial", alpha=0.1)</pre>
plot(fit, main=expression(paste(alpha,"=",0.1)))
par(op)
```

ncvsurv ncvsurv

Description

Fit coefficients paths for MCP- or SCAD-penalized Cox regression models over a grid of values for the regularization parameter lambda, with option for an additional L2 penalty.

Usage

```
ncvsurv(X, y, penalty=c("MCP", "SCAD", "lasso"),
gamma=switch(penalty, SCAD=3.7, 3), alpha=1,
lambda.min=ifelse(n>p,.001,.05), nlambda=100, lambda, eps=1e-4,
max.iter=10000, convex=TRUE, dfmax=p, penalty.factor=rep(1, ncol(X)),
warn=TRUE, returnX, ...)
```

Arguments

8	
X	The design matrix of predictor values. ncvsurv standardizes the data prior to fitting.
У	The time-to-event outcome, as a two-column matrix or Surv object. The first column should be time on study (follow up time); the second column should be a binary variable with 1 indicating that the event has occurred and 0 indicating (right) censoring.
penalty	The penalty to be applied to the model. Either "MCP" (the default), "SCAD", or "lasso".
gamma	The tuning parameter of the MCP/SCAD penalty (see details). Default is 3 for MCP and 3.7 for SCAD.
alpha	Tuning parameter for the Mnet estimator which controls the relative contributions from the MCP/SCAD penalty and the ridge, or L2 penalty. alpha=1 is equivalent to MCP/SCAD penalty, while alpha=0 would be equivalent to ridge regression. However, alpha=0 is not supported; alpha may be arbitrarily small, but not exactly 0.
lambda.min	The smallest value for lambda, as a fraction of lambda.max. Default is .001 if the number of observations is larger than the number of covariates and .05 otherwise.
nlambda	The number of lambda values. Default is 100.
lambda	A user-specified sequence of lambda values. By default, a sequence of values of length nlambda is computed, equally spaced on the log scale.
eps	Convergence threshhold. The algorithm iterates until the RMSD for the change in linear predictors for any coefficient is less than eps. Default is 1e-4.
max.iter	Maximum number of iterations (total across entire path). Default is 1000.
convex	Calculate index for which objective function ceases to be locally convex? Default is TRUE.
dfmax	Upper bound for the number of nonzero coefficients. Default is no upper bound. However, for large data sets, computational burden may be heavy for models with a large number of nonzero coefficients.

nevsury 19

penalty.factor A multiplicative factor for the penalty applied to each coefficient. If supplied,

penalty.factor must be a numeric vector of length equal to the number of columns of X. The purpose of penalty.factor is to apply differential penalization if some coefficients are thought to be more likely than others to be in the model. In particular, penalty.factor can be 0, in which case the coefficient is

always in the model without any penalization/shrinkage.

warn Return warning messages for failures to converge and model saturation? Default

is TRUE.

returnX Return the standardized design matrix along with the fit? By default, this option

is turned on if X is under 100 MB, but turned off for larger matrices to preserve memory. Note that certain methods, such as summary.ncvsurv require access

to the design matrix and may not be able to run if returnX=FALSE.

... Not used.

Details

The sequence of models indexed by the regularization parameter lambda is fit using a coordinate descent algorithm. In order to accomplish this, the second derivative (Hessian) of the Cox partial log-likelihood is diagonalized (see references for details). The objective function is defined to be

$$Q(\beta|X,y) = \frac{1}{n}L(\beta|X,y) + P_{\lambda}(\beta)$$

where the loss function L is the deviance (-2 times the partial log-likelihood) from the Cox regression mode. See here for more details.

Presently, ties are not handled by ncvsurv in a particularly sophisticated manner. This will be improved upon in a future release of ncvreg.

Value

An object with S3 class "ncvsurv" containing:

beta The fitted matrix of coefficients. The number of rows is equal to the number of coefficients, and the number of columns is equal to nlambda.

iter A vector of length nlambda containing the number of iterations until convergence at each value of lambda.

lambda The sequence of regularization parameter values in the path.

penalty Same as above.

model Same as above.

gamma Same as above.

alpha Same as above.

convex.min The last index for which the objective function is locally convex. The smallest value of lambda for which the objective function is convex is therefore lambda[convex.min], with corresponding coefficients beta[,convex.min].

loss The deviance of the fitted model at each value of lambda.

penalty.factor Same as above.

20 ncvsurv

n The number of observations.

For Cox models, the following objects are also returned (and are necessary to estimate baseline survival conditional on the estimated regression coefficients), all of which are ordered by time on study. I.e., the ith row of W does not correspond to the ith row of X):

W Matrix of exp(beta) values for each subject over all lambda values.

time Times on study.

fail Failure event indicator.

Additionally, if returnX=TRUE, the object will also contain

X The standardized design matrix.

Author(s)

Patrick Breheny

References

- Breheny P and Huang J. (2011) Coordinate descentalgorithms for nonconvex penalized regression, with applications to biological feature selection. *Annals of Applied Statistics*, **5**: 232-253. doi: 10.1214/10AOAS388
- Simon N, Friedman JH, Hastie T, and Tibshirani R. (2011) Regularization Paths for Cox's Proportional Hazards Model via Coordinate Descent. *Journal of Statistical Software*, **39**: 1-13. doi: 10.18637/jss.v039.i05

See Also

```
plot.ncvreg, cv.ncvsurv
```

Examples

```
data(Lung)
X <- Lung$X
y <- Lung$y
op \leftarrow par(mfrow=c(2,2))
fit <- ncvsurv(X, y)</pre>
plot(fit, main=expression(paste(gamma, "=", 3)))
fit <- ncvsurv(X, y, gamma=10)</pre>
plot(fit, main=expression(paste(gamma, "=",10)))
fit <- ncvsurv(X, y, gamma=1.5)</pre>
plot(fit, main=expression(paste(gamma, "=", 1.5)))
fit <- ncvsurv(X, y, penalty="SCAD")</pre>
plot(fit, main=expression(paste("SCAD, ",gamma,"=",3)))
par(op)
fit <- ncvsurv(X,y)</pre>
11 <- log(fit$lambda)</pre>
op <- par(mfrow=c(2,1))
```

perm.ncvreg 21

```
plot(11, BIC(fit), type="1", xlim=rev(range(11)))
lam <- fit$lambda[which.min(BIC(fit))]
b <- coef(fit, lambda=lam)
b[b!=0]
plot(fit)
abline(v=lam)
par(op)

S <- predict(fit, X, type='survival', lambda=lam)
plot(S, xlim=c(0,200))</pre>
```

perm.ncvreg

Permutation fitting for nevreg

Description

Fits multiple penalized regression models in which the outcome is randomly permuted, thereby allowing estimation of the marginal false discovery rate.

Usage

```
perm.ncvreg(X, y, ..., permute=c("outcome", "residuals"), N=10, seed, trace=FALSE)
```

Arguments

Χ	The design matrix, without an intercept, as in ncvreg.
У	The response vector, as in novreg.
• • •	Additional arguments to ncvreg.
permute	What to permute. If 'outcome', the response vector, y, is permuted. If 'residuals',

the residuals are permuted. This is only available for linear regression (i.e., for family='gaussian'). Note that permuting the residuals may take a long time, as the residuals differ for each value of lambda, so separate permutations are

required at every value of lambda. See also permres.

N The number of permutation replications. Default is 10.

seed You may set the seed of the random number generator in order to obtain repro-

ducible results.

trace If set to TRUE, perm.ncvreg will inform the user of its progress by announcing

the beginning of each permutation fit. Default is FALSE.

Details

The function fits a penalized regression model to the actual data, then repeats the process N times with a permuted version of the response vector. This allows estimation of the expected number of variables included by chance for each value of lambda. The ratio of this expected quantity to the number of selected variables using the actual (non-permuted) response is called the marginal false discovery rate (mFDR).

22 perm.ncvreg

Value

An object with S3 class "perm.ncvreg" containing:

EF The number of variables selected at each value of lambda, averaged over the

permutation fits.

S The actual number of selected variables for the non-permuted data.

mFDR The estimated marginal false discovery rate (EF/S).

fit The fitted novreg object for the original (non-permuted) data.

loss The loss/deviance for each value of lambda, averaged over the permutation fits.

This is an estimate of the explanatory power of the model under null conditions, and can be used to adjust the loss of the fitted model in a manner akin to the idea

of an adjusted R-squared in classical regression.

Author(s)

See Also

```
ncvreg, plot.mfdr, mfdr
```

Examples

```
# Linear regression ------
data(Prostate)
pmfit <- perm.ncvreg(Prostate$X, Prostate$y)</pre>
op \leftarrow par(mfcol=c(2,2))
plot(pmfit)
plot(pmfit, type="EF")
plot(pmfit$fit)
lam <- pmfit$fit$lambda</pre>
pmfit.r <- perm.ncvreg(Prostate$X, Prostate$y, permute='residuals')</pre>
plot(pmfit.r, col="red")
                                # Permuting residuals is
lines(lam, pmfit$mFDR, col="gray60") # less conservative
par(op)
# Logistic regression ------
data(Heart)
pmfit <- perm.ncvreg(Heart$X, Heart$y, family="binomial")</pre>
op <- par(mfcol=c(2,2))
plot(pmfit)
plot(pmfit, type="EF")
plot(pmfit$fit)
par(op)
```

permres 23

permres	Permute residuals for a fitted nevreg model	
per iiii es	i ermuie residuais jor a juied nevreg model	

Description

Fits multiple penalized regression models in which the residuals are randomly permuted, thereby allowing estimation of the marginal false discovery rate.

Usage

```
permres(fit, ...)
## S3 method for class 'ncvreg'
permres(fit, lambda, N=10, seed, trace=FALSE, ...)
```

Arguments

fit	A fitted nevreg model, as produced by nevreg(). To use with permres, the model must be fit using the returnX=TRUE option.
lambda	The regularization parameter to use for estimating residuals. Unlike perm.ncvreg, permres calculates EF and mFDR for a specific lambda value, not an entire path. As a result, it runs much faster.
N	The number of permutation replications. Default is 10.
seed	You may set the seed of the random number generator in order to obtain reproducible results.
trace	If set to TRUE, perm.ncvreg will inform the user of its progress by announcing the beginning of each permutation fit. Default is FALSE.
	Not used.

Details

The function fits a penalized regression model to the actual data, then repeats the process N times with a permuted version of the response vector. This allows estimation of the expected number of variables included by chance for each value of lambda. The ratio of this expected quantity to the number of selected variables using the actual (non-permuted) response is called the marginal false discovery rate (mFDR).

Value

A list with the following components:

EF	The number of variables selected at each value of lambda, averaged over the permutation fits.
S	The actual number of selected variables for the non-permuted data.
mFDR	The estimated marginal false discovery rate (EF/S).

24 plot.cv.ncvreg

loss

The loss/deviance, averaged over the permutation fits. This is an estimate of the explanatory power of the model under null conditions, and can be used to adjust the loss of the fitted model in a manner akin to the idea of an adjusted R-squared in classical regression.

Author(s)

Patrick Breheny patrick-breheny@uiowa.edu>

See Also

```
ncvreg, mfdr, perm.ncvreg
```

Examples

```
data(Prostate)
fit <- ncvreg(Prostate$X, Prostate$y, N=50)
permres(fit, lambda=0.15)</pre>
```

plot.cv.ncvreg

Plots the cross-validation curve from a cv.ncvreg object

Description

Plots the cross-validation curve from a cv.ncvreg or cv.ncvsurv object, along with standard error bars.

Usage

```
## S3 method for class 'cv.ncvreg'
plot(x, log.l=TRUE, type=c("cve", "rsq", "scale",
"snr", "pred", "all"), selected=TRUE, vertical.line=TRUE, col="red",
...)
```

Arguments

A cv.ncvreg or cv.ncvsurv object.

log.1 Should horizontal axis be on the log scale? Default is TRUE.

type What to plot on the vertical axis. cve plots the cross-validation error (deviance);

rsq plots an estimate of the fraction of the deviance explained by the model (R-squared); snr plots an estimate of the signal-to-noise ratio; scale plots, for family="gaussian", an estimate of the scale parameter (standard deviation); pred plots, for family="binomial", the estimated prediction error; all pro-

duces all of the above.

selected If TRUE (the default), places an axis on top of the plot denoting the number of

variables in the model (i.e., that have a nonzero regression coefficient) at that

value of lambda.

plot.cv.ncvreg 25

vertical.line	If TRUE (the default), draws a vertical line at the value where cross-validaton error is minimized.
col	Controls the color of the dots (CV estimates).
	Other graphical parameters to plot

Details

Error bars representing approximate 68% confidence intervals are plotted along with the estimates at value of lambda. For rsq and snr applied to models other than linear regression, the Cox-Snell R-squared is used.

Author(s)

Patrick Breheny

References

Breheny P and Huang J. (2011) Coordinate descentalgorithms for nonconvex penalized regression, with applications to biological feature selection. *Annals of Applied Statistics*, **5**: 232-253. doi: 10.1214/10AOAS388

See Also

```
ncvreg, cv.ncvreg
```

Examples

```
# Linear regression ------
data(Prostate)
cvfit <- cv.ncvreg(Prostate$X, Prostate$y)</pre>
plot(cvfit)
op <- par(mfrow=c(2,2))
plot(cvfit, type="all")
par(op)
# Logistic regression ------
data(Heart)
cvfit <- cv.ncvreg(Heart$X, Heart$y, family="binomial")</pre>
plot(cvfit)
op <- par(mfrow=c(2,2))
plot(cvfit, type="all")
par(op)
# Cox regression ------
data(Lung)
cvfit <- cv.ncvsurv(Lung$X, Lung$y)</pre>
op <- par(mfrow=c(1,2))</pre>
plot(cvfit)
plot(cvfit, type="rsq")
par(op)
```

26 plot.mfdr

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Plot marginal false discovery rate curves

Description

Plot marginal false discovery rate curves from an "mfdr" or "perm.ncvreg" object.

Usage

```
## S3 method for class 'mfdr'
plot(x, type=c("mFDR", "EF"), log.l=FALSE, selected=TRUE,
legend=TRUE, ...)
```

Arguments

x	A "perm.ncvreg" or "mfdr" object.
type	What to plot on the vertical axis. mFDR plots the marginal false discovery rate; EF plots the expected number of false discoveries along with the actual number of variables included in the model.
log.l	Should horizontal axis be on the log scale? Default is FALSE.
selected	If TRUE (the default), places an axis on top of the plot denoting the number of variables in the model (i.e., that have a nonzero regression coefficient) at that value of lambda.
legend	For type="EF" plots, draw a legend to indicate which line is for the actual selections and which line is for the expected number of false discoveries? Default is TRUE.
	Other graphical parameters to pass to plot

Author(s)

Patrick Breheny

References

Breheny P (2019). Marginal false discovery rates for penalized regression models. Biostatistics, 20: 299-314.

See Also

```
mfdr, perm.ncvreg
```

plot.ncvreg 27

Examples

```
data(Prostate)
fit <- ncvreg(Prostate$X, Prostate$y)</pre>
obj <- mfdr(fit)</pre>
obj[1:10,]
# Some plotting options
plot(obj)
plot(obj, type="EF")
plot(obj, log=TRUE)
# Comparison with perm.ncvreg
op <- par(mfrow=c(2,2))</pre>
plot(obj)
plot(obj, type="EF")
pmfit <- perm.ncvreg(Prostate$X, Prostate$y)</pre>
plot(pmfit)
plot(pmfit, type="EF")
par(op)
```

plot.ncvreg

Plot coefficients from a nevreg object

Description

Produces a plot of the coefficient paths for a fitted nevreg object.

Usage

```
## S3 method for class 'ncvreg'
plot(x, alpha=1, log.l=FALSE, shade=TRUE, col, ...)
```

Arguments

X	Fitted "ncvreg" model.
alpha	Controls alpha-blending, helpful when the number of covariates is large. Default is alpha=1.
log.l	Should horizontal axis be on the log scale? Default is FALSE.
shade	Should nonconvex region be shaded? Default is TRUE.
col	Vector of colors for coefficient lines. By default, evenly spaced colors are selected automatically.
	Other graphical parameters to plot

Author(s)

Patrick Breheny

28 plot.nevsurv.func

References

Breheny P and Huang J. (2011) Coordinate descentalgorithms for nonconvex penalized regression, with applications to biological feature selection. *Annals of Applied Statistics*, **5**: 232-253. doi: 10.1214/10AOAS388

See Also

ncvreg

Examples

```
data(Prostate)

fit <- ncvreg(Prostate$X, Prostate$y)
plot(fit)
plot(fit, col="black")
plot(fit, log=TRUE)</pre>
```

plot.ncvsurv.func

Plot survival curve for nevsurv model

Description

Plot survival curve for a model that has been fit using ncvsurv followed by a prediction of the survival function using predict.ncvsurv

Usage

```
## S3 method for class 'ncvsurv.func'
plot(x, alpha=1, ...)
```

Arguments

A 'ncvsurv.func' object, which is returned by predict.ncvsurv if type='survival' is specified. See examples.
 Controls alpha-blending (i.e., transparency). Useful if many overlapping lines are present.
 Other graphical parameters to pass to plot

Author(s)

Patrick Breheny

See Also

```
ncvsurv, predict.ncvsurv
```

predict.ncvreg 29

Examples

```
data(Lung)
X <- Lung$X
y <- Lung$y

fit <- ncvsurv(X, y)

# A single survival curve
S <- predict(fit, X[1,], type='survival', lambda=.15)
plot(S, xlim=c(0,200))

# Lots of survival curves
S <- predict(fit, X, type='survival', lambda=.08)
plot(S, xlim=c(0,200), alpha=0.3)</pre>
```

predict.ncvreg

Model predictions based on a fitted nevreg object.

Description

Similar to other predict methods, this function returns predictions from a fitted nevreg object.

Usage

```
## S3 method for class 'ncvreg'
predict(object, X, type=c("link", "response", "class",
"coefficients", "vars", "nvars"), lambda, which=1:length(object$lambda),
...)
## S3 method for class 'ncvreg'
coef(object, lambda, which=1:length(object$lambda),
drop=TRUE, ...)
```

Arguments

object	Fitted nevreg model object.
X	Matrix of values at which predictions are to be made. Not used for type="coefficients" or for some of the type settings in predict.
lambda	Values of the regularization parameter lambda at which predictions are requested. For values of lambda not in the sequence of fitted models, linear interpolation is used.
which	Indices of the penalty parameter lambda at which predictions are required. By default, all indices are returned. If lambda is specified, this will override which.
type	Type of prediction: "link" returns the linear predictors; "response" gives the fitted values; "class" returns the binomial outcome with the highest probability; "coefficients" returns the coefficients; "vars" returns a list containing the indices and names of the nonzero variables at each value of lambda; "nvars" returns the number of nonzero coefficients at each value of lambda.

30 predict.ncvsurv

drop If coefficients for a single value of lambda are to be returned, reduce dimensions to a vector? Setting drop=FALSE returns a 1-column matrix.

Not used.

Value

The object returned depends on type.

Author(s)

Patrick Breheny

References

Breheny P and Huang J. (2011) Coordinate descentalgorithms for nonconvex penalized regression, with applications to biological feature selection. *Annals of Applied Statistics*, **5**: 232-253. doi: 10.1214/10AOAS388

See Also

ncvreg

Examples

```
data(Heart)

fit <- ncvreg(Heart$X, Heart$y, family="binomial")
coef(fit, lambda=0.05)
head(predict(fit, Heart$X, type="link", lambda=0.05))
head(predict(fit, Heart$X, type="response", lambda=0.05))
head(predict(fit, Heart$X, type="class", lambda=0.05))
predict(fit, type="vars", lambda=c(0.05, 0.01))
predict(fit, type="nvars", lambda=c(0.05, 0.01))</pre>
```

predict.ncvsurv

Model predictions based on a fitted "ncvsurv" object.

Description

Similar to other predict methods, this function returns predictions from a fitted "ncvsurv" object.

Usage

```
## S3 method for class 'ncvsurv'
predict(object, X, type=c("link", "response", "survival",
"median", "coefficients", "vars", "nvars"), lambda,
which=1:length(object$lambda), ...)
```

31 predict.ncvsurv

Arguments

Fitted "ncvsurv" model object. object Matrix of values at which predictions are to be made. Not used for type="coefficients" or for some of the type settings in predict. lambda Values of the regularization parameter lambda at which predictions are requested. For values of lambda not in the sequence of fitted models, linear interpolation is used. which Indices of the penalty parameter lambda at which predictions are required. By default, all indices are returned. If lambda is specified, this will override which. Type of prediction: "link" returns the linear predictors; "response" gives type

the risk (i.e., exp(link)); "survival" returns the estimated survival function; "median" estimates median survival times. The other options are all identical to their nevreg counterparts: "coefficients" returns the coefficients; "vars" returns a list containing the indices and names of the nonzero variables at each value of lambda; "nvars" returns the number of nonzero coefficients at each value of lambda.

Not used. . . .

Details

Estimation of baseline survival function conditional on the estimated values of beta is carried out according to the method described in Chapter 4.3 of Kalbfleish and Prentice. In particular, it agrees exactly the results returned by survfit.coxph(...,type='kalbfleisch-prentice') in the survival package.

Value

The object returned depends on type.

Author(s)

Patrick Breheny patrick-breheny@uiowa.edu>

References

- Breheny P and Huang J. (2011) Coordinate descentalgorithms for nonconvex penalized regression, with applications to biological feature selection. Annals of Applied Statistics, 5: 232-253. doi: 10.1214/10AOAS388
- Kalbfleish JD and Prentice RL (2002). The Statistical Analysis of Failure Time Data, 2nd edition. Wiley.

See Also

ncvsurv

Prostate Prostate

Examples

```
data(Lung)
X <- Lung$X
y <- Lung$y
fit <- ncvsurv(X,y)</pre>
coef(fit, lambda=0.05)
head(predict(fit, X, type="link", lambda=0.05))
head(predict(fit, X, type="response", lambda=0.05))
# Survival function
S <- predict(fit, X[1,], type="survival", lambda=0.05)
S <- predict(fit, X, type="survival", lambda=0.05)</pre>
plot(S, xlim=c(0,200))
# Medians
predict(fit, X[1,], type="median", lambda=0.05)
M <- predict(fit, X, type="median")</pre>
M[1:10, 1:10]
# Nonzero coefficients
predict(fit, type="vars", lambda=c(0.1, 0.01))
predict(fit, type="nvars", lambda=c(0.1, 0.01))
```

Prostate

Factors associated with prostate specific antigen

Description

Data from a study by Stamey et al. (1989) to examine the association between prostate specific antigen (PSA) and several clinical measures that are potentially associated with PSA in men who were about to receive a radical prostatectomy. The variables are as follows:

- X: A design matrix with 97 instances (rows) and 8 predictor variables (columns). The columns are:
 - lcavol: Log cancer volume
 - lweight: Log prostate weight
 - age: The man's age
 - lbph: Log of the amount of benign hyperplasia
 - svi: Seminal vesicle invasion; 1=Yes, 0=No
 - lcp: Log of capsular penetration
 - gleason: Gleason score
 - pgg45: Percent of Gleason scores 4 or 5
- y: Log PSA

std 33

Usage

data(Prostate)

Source

https://web.stanford.edu/~hastie/ElemStatLearn/

References

- Hastie T, Tibshirani R, and Friedman J. (2001). The Elements of Statistical Learning. Springer.
- Stamey T, et al. (1989). Prostate specific antigen in the diagnosis and treatment of adenocarcinoma of the prostate. II. Radical prostatectomy treated patients. *Journal of Urology*, **16**: 1076-1083.

std

Standardizes a design matrix

Description

The function std accepts a design matrix and returns a standardized version of that matrix (i.e., each column will have mean 0 and mean sum of squares equal to 1).

Usage

std(X)

Arguments

Χ

A matrix (or object that can be coerced to a matrix, such as a data frame or numeric vector).

Details

This function centers and scales each column of X so that

$$\sum_{i=1}^{n} x_{ij} = 0$$

and

$$n^{-1} \sum_{i=1}^{n} x_{ij}^2 = 1$$

for all j. This is usually not necessary to call directly, as nowneg internally standardizes the design matrix, but inspection of the standardized design matrix can sometimes be useful. This differs from the base R function scale in two ways:

- 1. scale uses the sample standard deviation sqrt(sum(x^2)/(n-1)), while std uses the root-mean-square (population) standard deviation sqrt(mean(sum(x^2)))
- 2. std is faster.

34 summary.cv.ncvreg

Value

The standardized design matrix, with the following attribues:

- center, scale: mean and standard deviation used to scale the columns
- nonsingular: A vector indicating which columns of the original design matrix were able to be standardized (constant columns cannot be standardized to have a standard deviation of 1)

Examples

```
X <- matrix(rnorm(50), 10, 5)
S <- std(X)
apply(S, 2, sum)
apply(S, 2, function(x) mean(x^2))</pre>
```

summary.cv.ncvreg

Summarizing cross-validation-based inference

Description

Summary method for cv.ncvreg objects

Usage

```
## $3 method for class 'cv.ncvreg'
summary(object, ...)
## $3 method for class 'summary.cv.ncvreg'
print(x, digits, ...)
```

Arguments

object A "cv.ncvreg" or "cv.ncvsurv" object.

x A "summary.cv.ncvreg" object.

digits Number of digits past the decimal point to print out. Can be a vector specifying different display digits for each of the five non-integer printed values.

... Further arguments passed to or from other methods.

Value

summary.cv.ncvreg produces an object with S3 class "summary.cv.ncvreg". The class has its own print method and contains the following list elements:

```
penalty The penalty used by ncvreg.
```

model Either "linear" or "logistic", depending on the family option in novreg.

- **n** Number of observations
- **p** Number of regression coefficients (not including the intercept).

min The index of lambda with the smallest cross-validation error.

summary.ncvreg 35

lambda The sequence of lambda values used by cv.ncvreg.

cve Cross-validation error (deviance).

r.squared Proportion of variance explained by the model, as estimated by cross-validation. For models outside of linear regression, the Cox-Snell approach to defining R-squared is used.

snr Signal to noise ratio, as estimated by cross-validation.

sigma For linear regression models, the scale parameter estimate.

pe For logistic regression models, the prediction error (misclassification error).

Author(s)

Patrick Breheny

References

Breheny P and Huang J. (2011) Coordinate descentalgorithms for nonconvex penalized regression, with applications to biological feature selection. *Annals of Applied Statistics*, **5**: 232-253. doi: 10.1214/10AOAS388

See Also

```
ncvreg, cv.ncvreg, plot.cv.ncvreg
```

Examples

summary.ncvreg

Summary method for nevreg objects

Description

Inferential summaries for nevreg and nevsurv objects based on local marginal false discovery rates.

36 summary.ncvreg

Usage

```
## S3 method for class 'ncvreg'
summary(object, lambda, which, number, cutoff, ...)
## S3 method for class 'summary.ncvreg'
print(x, digits, ...)
```

Arguments

object An nevreg or nevsury object.

1 The regularization parameter value at which inference should be reported.

which Alternatively, lambda may be specified by index; which=10 means: report infer-

ence for the 10th value of lambda along the regularization path. If both lambda

and which are specified, lambda takes precedence.

number By default, summary will provide an inferential summary for each variable that

has been selected (i.e., each variable with a nonzero coefficient). Specifying number=5, for example, means that the summary table will include the 5 features with the lowest mfdr values, regardless of whether they were selected. To see

all features, number=Inf.

cutoff Alternatively, specifying for example cutoff=0.3 will report inference for all

features with mfdr under 30%. If both number and cutoff are specified, the

intersection between both sets of features is reported.

x A summary.ncvreg object.

digits Number of digits past the decimal point to print out. Can be a vector specifying

different display digits for each of the five non-integer printed values.

... Further arguments; in particular, if you have set returnX=FALSE, you will need

to supply X and y in order to calculate local mFDRs.

Value

summary.ncvreg and summary.ncvsurv produce object with S3 class summary.ncvreg. The class has its own print method and contains the following list elements:

penalty The penalty used by nowreg or nowsurv.

model Either "linear", "logistic", or "Cox".

n Number of instances.

p Number of regression coefficients (not including the intercept).

lambda The lambda value at which inference is being reported.

nvars The number of nonzero coefficients (again, not including the intercept) at that

value of lambda.

table A table containing estimates, normalized test statistics (z), and an estimate of

the local mfdr for each coefficient. The mfdr may be loosely interpreted, in an

empirical Bayes sense, as the probability that the given feature is null.

unpen.table If there are any unpenalized coefficients, a separate inferential summary is given

for them. Currently, this is based on lm/glm/coxph using the penalized coefficients to provide an offset. This is useful and more or less accurate, but not ideal; we hope to improve the inferential methods for unpenalized variables in

the future.

summary.ncvreg 37

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See Also

```
ncvreg, cv.ncvreg, plot.cv.ncvreg, local_mfdr
```

Examples

```
# Linear regression ------
data(Prostate)
fit <- ncvreg(Prostate$X, Prostate$y)</pre>
summary(fit, lambda=0.08)
# Logistic regression ------
data(Heart)
fit <- ncvreg(Heart$X, Heart$y, family="binomial")</pre>
summary(fit, lambda=0.05)
# Cox regression ------
data(Lung)
fit <- ncvsurv(Lung$X, Lung$y)</pre>
summary(fit, lambda=0.1)
# Options ------
fit <- ncvreg(Heart$X, Heart$y, family="binomial")</pre>
summary(fit, lambda=0.08, number=3)
summary(fit, lambda=0.08, number=Inf)
summary(fit, lambda=0.08, cutoff=0.5)
summary(fit, lambda=0.08, number=3, cutoff=0.5)
# If X and y are not returned with the fit, they must be supplied
fit <- ncvreg(Heart$X, Heart$y, family="binomial", returnX=FALSE)</pre>
summary(fit, X=Heart$X, y=Heart$y, lambda=0.08)
```

Index

```
* datasets
    Heart, 7
    Lung, 9
    Prostate, 32
AUC (AUC.cv.ncvsurv), 3
AUC.cv.ncvsurv, 3
coef.ncvreg(predict.ncvreg), 29
coef.ncvsurv(predict.ncvsurv), 30
cv.ncvreg, 4, 16, 25, 35, 37
cv.ncvsurv, 4, 20
cv.ncvsurv (cv.ncvreg), 4
fir, 6
Heart, 7
heart (Heart), 7
local_mfdr, 8, 37
Lung, 9
mfdr, 7, 10, 22, 24, 26
ncvfit. 12
ncvreg, 6, 11, 14, 22–25, 28, 30, 35, 37
ncvreg(), 8
ncvreg-package, 2
ncvsurv, 10, 11, 17, 28, 31
perm.ncvreg, 10, 11, 21, 23, 24, 26
permres, 21, 23
plot.cv.ncvreg, 6, 24, 35, 37
plot.mfdr, 11, 22, 26
plot.ncvreg, 16, 20, 27
plot.ncvsurv.func, 28
predict.ncvreg, 29
predict.ncvsurv, 28, 30
print.summary.cv.ncvreg
         (summary.cv.ncvreg), 34
```

```
print.summary.ncvreg (summary.ncvreg), 35
Prostate, 32
prostate (Prostate), 32
scale, 33
std, 33
summary.cv.ncvreg, 6, 34
summary.ncvreg, 15, 35
summary.ncvreg(), 8
summary.ncvsurv, 19
summary.ncvsurv (summary.ncvreg), 35
Surv, 18
survConcordance, 4
```