## Package 'spatialTIME'

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Title Spatial Analysis of Vectra Immunoflourescent Data

Version 1.2.1

**Description** Visualization and analysis of Vectra Immunoflourescent data. Options for calculating both the univariate and bivariate Ripley's K are included. Calculations are performed using a permutation-based approach presented by Wilson et al. <doi:10.1101/2021.04.27.21256104>.

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URL https://github.com/FridleyLab/spatialTIME

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#### NeedsCompilation no

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```
bi_NN_G
```

Bivariate Nearest Neighbor Based Measures of Spatial Clustering for IF data

#### Description

This function computes the nearest neighbor distribution for a particular marker relative to another marker for the observed and permuted point processes.

#### Usage

```
bi_NN_G(
 mif,
 mnames,
 r_range = seq(0, 100, 50),
 num_permutations = 50,
  edge_correction = "rs",
  keep_perm_dis = FALSE,
  exhaustive = TRUE,
 workers = 1,
  overwrite = FALSE,
 xloc = NULL,
 yloc = NULL
)
```

#### Arguments

mif	An MIF object
mnames	Character vector of marker names to estimate degree of nearest neighbor distribution
r_range	Numeric vector of potential r values this range must include 0. Note that the range selected is very different than count based measures. See details.

#### $bi_NN_G$

num_permutations		
	Numeric value indicating the number of permutations used. Default is 50.	
edge_correction	1	
	Character value indicating the type of edge correction to use. Options include "rs" or "hans".	
keep_perm_dis	Logical value determining whether or not to keep the full distribution of per- muted G values	
exhaustive	Logical. If TRUE then markers must be a vector and spatial measures will be computed all pairs of unique markers. If FALSE then markers must be a data.frame with the desired combinations.	
workers	Integer value for the number of workers to spawn	
overwrite	Logical value determining if you want the results to replace the current output (TRUE) or be to be appended (FALSE).	
xloc	a string corresponding to the $\boldsymbol{x}$ coordinates. If null the average of XMin and XMax will be used	
yloc	a string corresponding to the y coordinates. If null the average of YMin and YMax will be used	

#### Value

Returns a data frame

anchor	Marker for which the distances are measured from	
counted Theoretical CSR	Marker for which the distances are measured to	
	Expected value assuming complete spatial randomness	
Permuted CSR	Average observed G for the permuted point process	
Observed	Observed value for the observed point process	
Degree of Clustering Permuted		
	Degree of spatial clustering where the reference is the permuted estimate of CSR	
Degree of Clustering Theoretical		
	Degree of spatial clustering where the reference is the theoretical estimate of CSR	

#### Examples

```
#' #Create mif object
library(dplyr)
x <- create_mif(clinical_data = example_clinical %>%
mutate(deidentified_id = as.character(deidentified_id)),
sample_data = example_summary %>%
mutate(deidentified_id = as.character(deidentified_id)),
spatial_list = example_spatial,
patient_id = "deidentified_id",
sample_id = "deidentified_sample")
```

#Nearest Neighbor distribution for the colocalization of CD3+CD8+ positive

```
#cells and CD3+FOXP3+ positive cells where CD3+FOXP3+ is the reference cell
#type at neighborhood size of 10,20,...,100 (zero must be included in the
#input).
x <- bi_NN_G(mif = x, mnames = c("CD3..CD8.", "CD3..FOXP3."),
num_permutations = 1, edge_correction = 'rs', r = seq(0,100,10),</pre>
```

```
keep_perm_dis = FALSE, workers = 1, exhaustive = TRUE)
```

bi_ripleys_k	Bivariate Count Based Measures of Spatial Clustering function for IF
	data

#### Description

This function calculates count based Measures (Ripley's K, Besag L, and Marcon's M) of IF data to characterize correlation of the observed and permyted spatial point processes for two markers.

#### Usage

```
bi_ripleys_k(
  mif,
  mnames,
  r_range = seq(0, 100, 50),
  num_permutations = 50,
  edge_correction = "translation",
  method = "K",
  keep_perm_dis = FALSE,
  exhaustive = TRUE,
  workers = 1,
  overwrite = FALSE,
  xloc = NULL,
  yloc = NULL
)
```

#### Arguments

mif	An MIF object	
mnames	Character vector of marker names to estimate degree of spatial clustering. Spa- tial clustering will be computed between each combination of markers in this list.	
r_range	Numeric vector of potential r values this range must include 0	
num_permutations		
	Numeric value indicating the number of permutations used. Default is 50.	
edge_correction		
	Character value indicating the type of edge correction to use. Options include "theoretical", "translation", "isotropic" or "border". Various edges corrections are most appropriate in different settings. Default is "translation".	

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#### bi\_ripleys\_k

method	Character value indicating which measure (K, L, M) used to estimate the degree of spatial clustering. Description of the methods can be found in Details section.
keep_perm_dis	Logical value determining whether or not to keep the full distribution of per- muted K values
exhaustive	Logical. If TRUE then markers must be a vector and spatial measures will be computed all pairs of unique markers. If FALSE then markers must be a data.frame with the desired combinations.
workers	Integer value for the number of workers to spawn
overwrite	Logical value determining if you want the results to replace the current output (TRUE) or be to be appended (FALSE).
xloc	a string corresponding to the x coordinates. If null the average of XMin and XMax will be used
yloc	a string corresponding to the y coordinates. If null the average of YMin and YMax will be used

#### Value

Returns a data frame

anchor	Marker for which the distances are measured from	
counted Theoretical CSR	Marker for which the distances are measured to	
	Expected value assuming complete spatial randomness	
Permuted CSR	Average observed K, L, or M for the permuted point process	
Observed	Observed value for the observed point process	
Degree of Clustering Permuted		
	Degree of spatial clustering where the reference is the permuted estimate of CSR	
Degree of Clustering Theoretical		
	Degree of spatial clustering where the reference is the theoretical estimate of CSR	

#### Examples

```
#' #Create mif object
library(dplyr)
x <- create_mif(clinical_data = example_clinical %>%
mutate(deidentified_id = as.character(deidentified_id)),
sample_data = example_summary %>%
mutate(deidentified_id = as.character(deidentified_id)),
spatial_list = example_spatial,
patient_id = "deidentified_id",
sample_id = "deidentified_sample")
```

```
#Ripley's K for the colocalization of CD3+CD8+ positive cells and
#CD3+FOXP3+ positive cells where CD3+FOXP3+ is the reference cell type at
#neighborhood size of 10,20,...,100 (zero must be included in the input).
```

```
x <- bi_ripleys_k(mif = x, mnames = c("CD3..CD8.", "CD3..FOXP3."),
num_permutations = 1, edge_correction = 'translation', r = seq(0,100,10),
keep_perm_dis = FALSE, workers = 1, exhaustive = TRUE)
```

create\_mif

Create Multiplex Immunoflourescent object

#### Description

Creates an MIF object for use in spatialIF functions

#### Usage

```
create_mif(
  clinical_data,
  sample_data,
  spatial_list = NULL,
  patient_id = "patient_id",
  sample_id = "image_tag"
)
```

#### Arguments

clinical_data	A data frame containing patient level data with one row per participant.
sample_data	A data frame containing sample level data with one row per sample. Should at a minimum contain a 2 columns: one for sample names and one for the corresponding patient name.
spatial_list	A named list of data frames with the spatial data from each sample making up each individual data frame
patient_id	A character string indicating the column name for patient id in sample and clin- ical data frames.
sample_id	A character string indicating the column name for sample id in the sample data frame

#### Value

Returns a custom MIF

clinical	Data frame of clinical data
sample	Data frame of sample data
spatial	Named list of spatial data
derived	List of data derived using the MIF object
patient_id	The column name for sample id in the sample data frame with the clinical data
sample_id	The column name for sample id in the sample data frame to merge with the spatial data

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#### example\_clinical

#### Examples

```
#Create mif object
library(dplyr)
x <- create_mif(clinical_data = example_clinical %>%
mutate(deidentified_id = as.character(deidentified_id)),
sample_data = example_summary %>%
mutate(deidentified_id = as.character(deidentified_id)),
spatial_list = example_spatial,
patient_id = "deidentified_id",
sample_id = "deidentified_sample")
```

example\_clinical Clinical variables of 229 patients

#### Description

A tibble wuith clinical characteristics for 229 patients

#### Usage

example\_clinical

#### Format

A tibble with 229 rows and 6 variables

**age** age at diagnosis

race self-idenitifed race

sex patient biological sex

status disease status

deidenitifed\_sample sample identifier

deidentified\_id patient identifier

example\_spatial Example list of 5 spatial TMA data

#### Description

A list containing 5 spatial data frames

#### Usage

example\_spatial

#### Format

A list of 5 data frames:

- TMA\_[3,B].tiff
- TMA\_[6,F].tiff
- TMA\_[7,B].tiff
- TMA\_[9,K].tiff
- TMA\_[8,U].tiff

example\_summary Marker summaries of 229 samples

#### Description

A dataset containing summaries of 25 markers and 229 samples

#### Usage

```
example_summary
```

#### Format

A tibble with 229 rows and 29 variables:

deidentified\_id patient-level id

deidentified\_sample sample-level id ...

NN\_G

Nearest Neighbor Based Measures of Spatial Clustering for IF data

#### Description

For a given cell type, this function computes proportion of cells that have nearest neighbor less than r for the observed and permuted point processes.

#### $NN_G$

#### Usage

```
NN_G(
   mif,
   mnames,
   r_range = seq(0, 100, 50),
   num_permutations = 50,
   edge_correction = "rs",
   keep_perm_dis = FALSE,
   workers = 1,
   overwrite = FALSE,
   xloc = NULL,
   yloc = NULL
)
```

#### Arguments

mif	An MIF object
mnames	Character vector of marker names to estimate degree of nearest neighbor distribution
r_range	Numeric vector of potential r values this range must include 0.
num_permutation	ns
	Numeric value indicating the number of permutations used. Default is 50.
edge_correction	n
	Character value indicating the type of edge correction to use. Options include "rs" or "hans".
keep_perm_dis	Logical value determining whether or not to keep the full distribution of per- muted G values
workers	Integer value for the number of workers to spawn
overwrite	Logical value determining if you want the results to replace the current output (TRUE) or be to be appended (FALSE).
xloc	a string corresponding to the x coordinates. If null the average of XMin and XMax will be used
yloc	a string corresponding to the y coordinates. If null the average of YMin and YMax will be used

#### Value

Returns a data.frame	
Theoretical CSR	
	Expected value assuming complete spatial randomnessn
Permuted CSR	Average observed G for the permuted point process
Observed	Observed value for the observed point process
Degree of Clustering Permuted	
	Degree of spatial clustering where the reference is the permuted estimate of CSR

Degree of Clustering Theoretical

Degree of spatial clustering where the reference is the theoretical estimate of CSR

#### Examples

```
#Create mif object
library(dplyr)
x <- create_mif(clinical_data = example_clinical %>%
mutate(deidentified_id = as.character(deidentified_id)),
sample_data = example_summary %>%
mutate(deidentified_id = as.character(deidentified_id)),
spatial_list = example_spatial,
patient_id = "deidentified_id",
sample_id = "deidentified_sample")
# Define the set of markers to study
markers <- c("CD3..0pal.570..Positive","CD8..0pal.520..Positive",</pre>
"FOXP3..Opal.620..Positive", "CD3..CD8.", "CD3..FOXP3.")
# Nearest Neighbor distribution for all markers with a neighborhood size
# of 10,20,...,100 (zero must be included in the input).
x <- NN_G(mif = x, mnames = markers, num_permutations = 1,</pre>
edge_correction = 'rs', r = seq(0,100,10),
keep_perm_dis = FALSE, workers = 1)
```

plot_immunoflo	Generate plot of TMA po	oint process

#### Description

This function generates plot of point process in rectangular or circular window.

#### Usage

```
plot_immunoflo(
  mif,
  plot_title,
  mnames,
  mcolors = NULL,
  cell_type = NULL,
  filename = NULL,
  path = NULL
)
```

#### ripleys\_k

#### Arguments

mif	MIF object created using create_MIF().
plot_title	Character string or vector of character strings of variable name(s) to serve as plot title(s).
mnames	Character vector containing marker names.
mcolors	Character vector of color names to display markers in the plot.
cell_type	Character vector of cell type
filename	Character string of file name to store plots. Plots are generated as single .pdf file.
path	Different path than file name or to use in conjunction with filename ???

#### Value

mif object and the ggplot objects can be viewed form the derived slot of the mif object

#### Examples

```
#Create mif object
library(dplyr)
x <- create_mif(clinical_data = example_clinical %>%
mutate(deidentified_id = as.character(deidentified_id)),
sample_data = example_summary %>%
mutate(deidentified_id = as.character(deidentified_id)),
spatial_list = example_spatial,
patient_id = "deidentified_id",
sample_id = "deidentified_sample")
mnames_good <- c("CD3..Opal.570..Positive", "CD8..Opal.520..Positive",
"FOXP3..Opal.620..Positive", "PDL1..Opal.540..Positive",
"FOXP3..Opal.620..Positive", "CD3..CD8.", "CD3..FOXP3.")
x <- plot_immunoflo(x, plot_title = "deidentified_sample", mnames = mnames_good,
cell_type = "Classifier.Label")
x[["derived"]][["spatial_plots"]][[4]]
```

ripleys\_k

Calculate Count Based Measures of Spatial Clustering for IF data

#### Description

This function calculates count based Measures (Ripley's K, Besag L, and Marcon's M) of IF data to characterize correlation of spatial point process.

#### Usage

```
ripleys_k(
  mif,
  mnames,
  r_range = seq(0, 100, 50),
  num_permutations = 50,
  edge_correction = "translation",
  method = "K",
  keep_perm_dis = FALSE,
  workers = 1,
  overwrite = FALSE,
  xloc = NULL,
  yloc = NULL
)
```

#### Arguments

mif	An MIF object
mnames	Character vector of marker names to estimate degree of spatial clustering.
r_range	Numeric vector of potential r values this range must include 0.
num_permutatio	ns
	Numeric value indicating the number of permutations used. Default is 50.
edge_correctio	n
	Character value indicating the type of edge correction to use. Options include "translation" or "isotropic".
method	Character value indicating which measure (K, L, M) used to estimate the degree of spatial clustering. Description of the methods can be found in Details section.
keep_perm_dis	Logical value determining whether or not to keep the full distribution of per- muted K values
workers	Integer value for the number of workers to spawn
overwrite	Logical value determining if you want the results to replace the current output (TRUE) or be to be appended (FALSE).
xloc	a string corresponding to the x coordinates. If null the average of XMin and XMax will be used
yloc	a string corresponding to the y coordinates. If null the average of YMin and YMax will be used

#### Value

Returns a data.frame

Theoretical CSR	
	Expected value assuming complete spatial randomnessn
Permuted CSR	Average observed K, L, or M for the permuted point process
Observed	Observed value for the observed point process

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Degree of Clustering Permuted Degree of spatial clustering where the reference is the permutated estimate of CSR Degree of Clustering Theoretical Degree of spatial clustering where the reference is the theoretical estimate of CSR

#### Examples

```
#Create mif object
library(dplyr)
x <- create_mif(clinical_data = example_clinical %>%
mutate(deidentified_id = as.character(deidentified_id)),
sample_data = example_summary %>%
mutate(deidentified_id = as.character(deidentified_id)),
spatial_list = example_spatial,
patient_id = "deidentified_id",
sample_id = "deidentified_sample")
# Define the set of markers to study
markers <- c("CD3..0pal.570..Positive","CD8..0pal.520..Positive",</pre>
"FOXP3..Opal.620..Positive", "CD3..CD8.", "CD3..FOXP3.")
# Ripley's K for all markers with a neighborhood size
# of 10,20,...,100 (zero must be included in the input).
x <- ripleys_k(mif = x, mnames = markers, num_permutations = 1,</pre>
edge_correction = 'translation', r = seq(0,100,10),
```

```
keep_perm_dis = FALSE, workers = 1)
```

subset\_mif

Subset mif object on cellular level

#### Description

This function allows to subset the mif object into compartments. For instance a mif object includes all cells and the desired analysis is based on only the tumor or stroma compartment then this function will subset the spatial list to just the cells in the desired compartment

#### Usage

```
subset_mif(mif, classifier, level, markers)
```

#### Arguments

mif	An MIF object
classifier	Column name for spatial dataframe to subset
level	Determines which level of the classifier to keep.
markers	vector of

#### Value

mif object where the spatial list only as the cell that are the specified level.

#### Examples

```
#' #Create mif object
library(dplyr)
x <- create_mif(clinical_data = example_clinical %>%
mutate(deidentified_id = as.character(deidentified_id)),
sample_data = example_summary %>%
mutate(deidentified_id = as.character(deidentified_id)),
spatial_list = example_spatial,
patient_id = "deidentified_id",
sample_id = "deidentified_sample")
markers = c("CD3..0pal.570..Positive", "CD8..0pal.520..Positive",
"FOXP3..0pal.620..Positive", "PD11..0pal.540..Positive",
"PD1..0pal.650..Positive", "CD3..CD8.", "CD3..FOXP3.")
```

level = 'Tumor', markers = markers)

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